



Published in final edited form as:

Respir Physiol Neurobiol. 2019 December ; 270: 103259. doi:10.1016/j.resp.2019.103259.

Respiratory rhythm generation, hypoxia, and oxidative stress— Implications for development

Alfredo J. Garcia III^a, Jean Charles Viemari^b, Maggie A. Khuu^a

^aInstitute for Integrative Physiology, Section of Emergency Medicine, The University of Chicago, Chicago, 60637, IL, United States

^bInstitut de Neurosciences de la Timone, P3M team, UMR7289 CNRS & AMU, Faculté de Médecine de la Timone, 27 Bd Jean Moulin, Marseille, 13005, France

Abstract

Encountered in a number of clinical conditions, repeated hypoxia/reoxygenation during the neonatal period can pose both a threat to immediate survival as well as a diminished quality of living later in life. This review focuses on our current understanding of central respiratory rhythm generation and the role that hypoxia and reoxygenation play in influencing rhythmogenesis. Here, we examine the stereotypical response of the inspiratory rhythm from the preBötzing complex (preBötC), basic neuronal mechanisms that support rhythm generation during the peri-hypoxic interval, and the physiological consequences of inspiratory network responsiveness to hypoxia and reoxygenation, acute and chronic intermittent hypoxia, and oxidative stress. These topics are examined in the context of Sudden Infant Death Syndrome, apneas of prematurity, and neonatal abstinence syndrome.

1. Introduction

The challenge of ventilation begins with our first breath. Immediately following birth, the transition from the low oxygen intrauterine environment to the oxygen rich state of room air becomes a significant life-threatening challenge when stable breathing is not yet established (Alvaro and Rigatto, 2016; Hillman et al., 2012). Maternal issues, such as maternal hypertension, preeclampsia (Hubbard and Shingleton, 1985) and gestational diabetes (Gross et al., 1983) can lead to premature birth, which can result in the instability of neonatal breathing. When combined with immediate birthing complications, such as aspiration of meconium-stained amniotic fluids, caesarian-section delivery, and neonatal abstinence syndrome, the risk for respiratory instability increases substantially (“Neonatal drug withdrawal. American Academy of Pediatrics Committee on Drugs,” 1998; Reuter et al., 2014). Unstable breathing during the neonatal period can quickly transition from a couple of skipped breaths to a prolonged hypoxic insult, which can result in life-long problems that extend far beyond breathing, and in the most extreme cases, cause infant death (Bancalari and Claire, 2018). Even after the immediate perinatal period, during the first weeks of life, infants, that otherwise appear healthy, are at risk for Sudden Infant Death Syndrome (SIDS), an outcome where oscillations in blood oxygenation is a factor that can lead to SIDS (Waggener et al., 1990). The gravity of such outcomes due to respiratory instability are clinically well-recognized. Thus, when the potential for unstable neonatal breathing arises,

practitioners place significant effort to monitor neonatal breathing and mitigate the occurrence of hypoxia. However, since our understanding into the basis and development of respiratory control in the neonate remains incomplete, there are few options to treat unstable breathing in the newborn. This review examines current advances made in understanding the development and involvement of brainstem networks in controlling respiration in response to and recovery from hypoxia. We will focus particularly on findings derived from the study of inspiratory rhythm generation from the preBötzinger complex (preBötC) during different forms of hypoxia and exposure to reactive oxygen species.

1.1. The neonatal ventilatory response to hypoxia and the perinatal respiratory networks of the brainstem

Respiratory control can be broadly categorized as sensory input, rhythm generation, and motor output (Barlow and Estep, 2006; Bellingham, 1998; Braman, 1995; Grillner, 2003). Balanced integration across these areas of control is necessary for maintaining stable and dynamically responsive breathing (Estelle B. Gauda and Martin, 2012). Yet in the neonate, many aspects of respiratory control are still developing. In rodents, the adult ventilatory response to hypercapnia appears to plateau following first week of life and into adulthood (Stunden et al., 2001) and similarly, carotid body discharge in response to hypoxia, hypercapnia, or hypoxic hypercapnia is stronger in adult (> 5 weeks of age) when compared to the neonate (P5 to P7 of age) (Pepper et al., 1995). Thus, it is during the neonatal period that susceptibility to experiencing a hypoxic event may be enhanced due to underdeveloped responses to deviations on blood gases.

Term and preterm (< 38 week) human infants exhibit a biphasic ventilatory response to drops in oxygen (Martin et al., 1998; Rigatto et al., 1975). It is characterized by an initial rapid, yet transient, increase in minute ventilation. This initial increase is later followed by a decline in ventilation that often falls well-below baseline breathing. This stereotypical ventilatory response to hypoxia is preserved also in neonatal rodents (Garcia et al., 2017), which have served as useful models for studying brainstem networks involved with the control of breathing and the mechanisms that support the responsiveness of neonatal breathing to hypoxia (Garcia et al., 2017; Viemari et al., 2003).

The rodent respiratory network is distributed throughout the brainstem and is composed of at least three groups of network oscillators: the preBötzinger complex (preBötC)(Anderson and Ramirez, 2017; Del Negro et al., 2018); the retrotrapezoid/parafacial respiratory group (RTN/pFRG)(Guyenet and Mulkey, 2010), and the recently discovered post-inspiratory complex (PiCo) (Anderson et al., 2016). While PiCo has been described in neonate (Anderson et al., 2016), it is unknown whether this network is active in the embryo. However, both the preBötC and RTN/pFRG appear to be active well before birth(Guyenet and Mulkey, 2010; Thoby-Brisson et al., 2005). Inspiratory activity from the brainstem can be recorded from the spinal cord respiratory motor pools as early as embryonic day 16 (E16) (Viemari et al., 2003). Respiratory activity from the embryonic brainstem emerges as a result of interactions between the preBötC and the RTN/pFRG (Champagnat et al., 2011; Tupal et al., 2014). While rhythmicity from the preBötC starts between E15 to E18 (Chevalier et al., 2016a,b; Pagliardini et al., 2003; Thoby-Brisson et al., 2005), activity from RTN/pFRG is

present at E14.5 (Thoby-Brisson et al., 2009). During the early postnatal development, RTN/pFRG undergoes a shift in activity from a pre-inspiratory to an inspiratory pattern while preBötC activity remained unchanged (Oku et al., 2007). In juveniles (postnatal day 7 to 13) transection below the RTN/pFRG abolishes active expiration but does not affect inspiration (Janczewski and Feldman, 2006). Thus, while the preBötC is responsible for the inspiratory rhythm, the RTN/pFRG forms the basis of a functionally independent expiratory oscillator (Thoby-Brisson et al., 2009). The RTN/pFRG also contains several cellular constituents such as Phox2b expressing neurons that provide chemosensory drive in response to CO₂ and acidification (Guyenet et al., 2005; Stornetta et al., 2006) and astrocytes that contribute to central chemoreception (Mulkey and Wenker, 2011; Wenker et al., 2010). Inspiratory activity at embryonic ages exhibits sensitivity to neuromodulation (Di Pasquale et al., 1994; Fujii and Arata, 2010; Viemari et al., 2003) and can respond to changes in oxygenation (Chevalier et al., 2016a,b; Viemari et al., 2003) indicating that the respiratory network has the potential to be dynamically responsive well-before birth. However, many cellular and physiological aspects of respiratory control continue to develop and change well-after birth (Prabhakar et al., 2007; Stunden et al., 2001; Wong-Riley and Liu, 2005) which leaves the neonate without the full complement of mechanisms to support the stability of adult breathing and may create vulnerabilities during the neonatal period where the responsiveness of breathing is underdeveloped (Rigatto and Brady, 1972).

1.2. Fundamental elements of inspiratory rhythm generation

The preBötC can be isolated in a single brainstem slice preparation and is where the principles and mechanisms underpinning inspiratory rhythmogenesis have been extensively studied (Butera et al., 1999; Koshiya and Smith, 1999; Smith et al., 1991). In the seminal work localizing the preBötC by Smith et al. (Smith et al., 1991), rhythm generation from the inspiratory network was shown to be attenuated in response to microinjection of the AMPA receptor antagonist, CNQX, suggesting that inspiratory rhythmogenesis from the preBötC is largely dependent on excitatory transmission. Subsequent studies examining the basis of rhythm generation in the preBötC have confirmed this observation (Lieske and Ramirez, 2006; Morgado-Valle and Feldman, 2007; Shao et al., 2003) and the necessity of glutamatergic transmission for rhythm generation is a well-accepted key principle. In isolation of fast synaptic transmission, individual preBötC neurons can be categorized as intrinsic bursting neurons, weak/tonic spiking neurons, or silent neurons (Ramirez et al., 2011). Intrinsic bursting neurons in the inspiratory network can be further divided by the underlying conductances that drive bursting into either persistent sodium current (I_{NaP}) bursters or non-specific cation current (I_{CAN}) bursters (Ramirez et al., 2004). Additional intrinsic and synaptic properties contribute to stabilizing and shaping the phenotype of these general preBötC classes of preBötC neurons, and we refer the reader to the following reviews (Ramirez et al., 2012, 2011; Ramirez et al., 2004) for a more extensive understanding of the cellular and network properties that support rhythmogenesis from the preBötC beyond the peri-hypoxic interval.

1.3. Oxygenation and the preBötC

The isolated preBötC is capable of generating different neural rhythms reminiscent of eupnea, gasping, and sighing (Lieske et al., 2000) and can be revealed by changing the state

of oxygenation surrounding the network (Garcia et al., 2017, 2013b; Hill et al., 2011; Lieske et al., 2000). When the inspiratory network becomes initially hypoxic, normal rhythmogenesis (Fig. 1A) increases in frequency and begins to generate augmented bursts (Fig. 1B) while later in hypoxia, augmented bursting ceases and rhythmogenesis slows (Fig. 1C). This initial augmentation followed by depression is remarkably similar to the neonatal ventilatory response to hypoxia (Garcia et al., 2016; Pena et al., 2008). Upon reoxygenation, a paradoxical depression in in vitro rhythmogenesis occurs (Fig. 1D) which is also strikingly similar to post hypoxic ventilatory depression (Coles et al., 1998; Garcia et al., 2013b). The similarities between the stereotypical in vitro response of the rhythm generating network to hypoxia and the stereotypical hypoxic response of neonatal breathing supports the notion that the preBötC is a significant contributor to all phases of breathing during the peri-hypoxic interval.

Although it is tempting to conclude that the preBötC is responsible for driving changes in the breathing rhythm during transitions in oxygenation, rhythmogenesis from the preBötC appears to stereotypically respond to relative changes in oxygenation as opposed to sensing absolute changes in the oxygen environment (Hill et al., 2011). This observation suggests that the contribution of the preBötC is only responsive to changes in oxygenation and sensory input from outside inspiratory network is still required for normal adult physiology. Indeed, targeted microinjections of colchicine to block axonal transport in the adult preBötC reduces, but does not eliminate, both hypercapnic and hypoxic ventilatory responses (Wu et al., 2005). During the neonatal period when central CO_2/H^+ chemoreception (Cerpa et al., 2017; Stunden et al., 2001), peripheral O_2 (Pawar et al., 2008) and peripheral CO_2/H^+ (Pepper et al., 1995) chemoreception are still developing, the intrinsic responsiveness of the preBötC to changes in oxygenation may have a greater role in influencing neonatal breathing responses to deviations in blood gases.

1.4. Neuromodulation of rhythmogenesis during hypoxia

Intrinsic properties of individual preBötC neurons play an important role in maintaining hypoxic rhythmogenesis. In particular, I_{NaP} -dependent activity appears to be critical for driving the preBötC rhythm under low O_2 tensions as riluzole and R56865, I_{NaP} blockers, prevent hypoxic rhythmogenesis, but has little effect on rhythmogenesis under well-oxygenated control conditions (Paton et al., 2006; Pena et al., 2004). Intracellular recordings from synaptically isolated preBötC neurons also demonstrate that I_{NaP} dependent bursters are intrinsically active during hypoxia while I_{CAN} and weak/tonic spiking neurons are silenced by hypoxia (Pena et al., 2004).

Under well-oxygenated conditions, the majority of preBötC neurons normally exist as weak/tonic spiking neurons while silent and intrinsic bursting neurons only constitute a minority (Carroll and Ramirez, 2013; Carroll et al., 2013; Nieto-Posadas et al., 2014). This composition provides sufficient stability in well-oxygenated states, yet in the presence of enhanced serotonergic (Tryba et al., 2006), substance P-ergic (Pena and Ramirez, 2004), or adrenergic (Viemari and Ramirez, 2006) neuromodulation, weak/tonic spiking neurons can assume a burster phenotype. These observations show that firing phenotype is not a fixed property among all preBötC neurons. Increasing intrinsically bursting neurons throughout

the respiratory network may enhance the robustness of rhythm generation when the inspiratory network is burdened. Recent multielectrode studies examining network interaction during hypoxic rhythmogenesis indicate that during the hypoxia, functional connectivity appears to wane when compared to rhythmogenesis under normal conditions (Nieto-Posadas et al., 2014). However, when serotonergic 2A (Pena and Ramirez, 2002) or alpha-2 adrenergic receptors (Viemari et al., 2011) are blocked during a transition to hypoxia, the loss of neuromodulatory signaling increases the likelihood for failed rhythmogenesis.

in vivo exposure to intermittent hypoxia leads to an increase in catecholaminergic neurons of the ventral lateral medulla and in gene expression of serotonin transporter (Slc6a4) 5-HTT and tryptophan hydroxylase 2, two critical components important to maintenance of serotonergic neuromodulation (Givan and Cummings, 2016). Furthermore, loss of central serotonergic neuromodulation increases the chance of failed autoresuscitation (Chen et al., 2013), impairs hypoxia-triggered arousal (Young et al., 2017), and increases apneas (Hodges et al., 2009; Young et al., 2017). Combined with the observations made in the inspiratory network, these findings suggest that enhancing neuromodulation is a natural response to the recurrence of hypoxia and may serve as a potential mechanism that defends against unstable breathing by supporting reliable rhythmogenesis in response to changing blood gases.

1.5. SIDS, hypoxia, and post-hypoxic recovery

Sudden infant death syndrome (SIDS) is a diagnosis provided when the death of an infant less than one year of age cannot be explained by other means (Byard, 2018; Ferguson, 2015). The triple-risk model of SIDS first proposed by Filliano and Kinney (Filliano and Kinney, 1994) states that the SIDS emerges as the result of intersection among three factors: a vulnerable infant, a critical developmental period, and an exogenous stressor (i.e., environmental challenge). Biological sex and age are thought to be significant factors that contribute to the first two factors in the triple-risk model of SIDS. Males are more likely to succumb to SIDS (Mage and Donner, 2004, 2014) while the peak occurrence of SIDS is between three and six months (Mage and Donner, 2009). Furthermore, postmortem analysis suggests that deficits in serotonergic neuromodulation may further increase vulnerability to SIDS (Kinney et al., 2001). Thus, if a vulnerable infant encounters even the briefest hypoxic challenge, the threshold for SIDS may be much lower (Ramirez et al., 2018; Thach, 2008). In the sleeping infant, the prone position increases likelihood for unescapable hypoxic exposure and the susceptibility for suffocation due to the potential inability of the infant in repositioning his or her head to prevent rebreathing into the cushion, pillow, or bedding (Erck Lambert et al., 2019). Rebreathing into cushion, pillow, or bedding, will initially lead to hypercapnia and subsequently, if left unchecked hypoxia. Impaired central and peripheral chemoreception have been largely recognized as significant contributing factors to SIDS occurrence (For review see: (E. B. Gauda et al., 2007; Porzionato et al., 2018)). For example, hyposensitivity of peripheral chemoreception may contribute not only to the blunted ventilatory response to hypoxic hypercapnia, but also may lead to a failure in arousal. Thus, deficits in the arousal response during these hypoxic challenges are thought to contribute to these outcomes (Garcia et al., 2013a). Together hypoxia and the failure to arouse to such challenge may be considered two significant factors contributing to SIDS.

Further supporting the perspective that hypoxic challenge is a significant factor that can lead to infant death, increased rates of both SIDS (Barkin et al., 1981) and unexplained neonatal hospital death (Yangzom et al., 2008) have been documented in high altitude populations. Hypoxia may alter neonatal cardiac maturation, leading to the impaired cardiac conduction, and decreasing the threshold for cardiac arrhythmias and the occurrence of sudden death (Neary et al., 2013). In cases of prematurity, hypoxia may result from underdevelopment of respiratory centers that cause hypoventilation and diaphragmatic malfunction. Other studies have also demonstrated that intrauterine growth restriction via prenatal hypoxia not only leads to a modification in inspiratory rhythm but also alterations in catecholaminergic modulation (Tree et al., 2016). However, recent findings suggest the period of post-hypoxic recovery of breathing and rhythmogenesis from the preBötC may also be a factor contributing to SIDS.

Following hypoxia, rhythmogenesis is initially suppressed but later frequency rebounds (Garcia et al., 2013b; Hill et al., 2011). This biphasic response of preBötC rhythm may contribute to the paradoxical occurrence of post-hypoxic ventilatory depression—a transition period that can increase the chance for unstable breathing and contribute to the risk of SIDS. Both postnatal age and sex appear to be factors that can influence inspiratory rhythmogenesis during reoxygenation (Garcia et al., 2013b). The latency of post-hypoxic recovery in rhythmogenesis from the preBötC increases with postnatal age (Garcia et al., 2013b). During the second week of life, NMDA receptor expression (Liu and Wong-Riley, 2005), GABA_A receptor subunit expression (Liu and Wong-Riley, 2006) and chloride homeostasis (Liu and Wong-Riley, 2012) change and may contribute to the prolongation of post-hypoxic recovery. In addition to the prolongation of post-hypoxic recovery, a sex-based difference emerges in the post-hypoxic recovery of preBötC rhythms during the second week of life (Garcia et al., 2013b) whereby post hypoxic depression is prolonged in the preBötC rhythms from males when compared to females. This sex-based difference in post-hypoxic recovery was eliminated by either activating or blocking the ATP-sensitive potassium channel (K_{ATP}), suggesting that the dynamic activity of the K_{ATP} largely contributes to the post-hypoxic recovery of the rhythmogenesis from the preBötC as opposed to other potential differences such as expression of the channel. Thus, as the activity of K_{ATP} is regulated by the availability of ATP, sex-based differences in metabolic activity within the respiratory network may impact the breathing rhythm during peri-hypoxic intervals.

1.6. Reactive oxygen species

As discussed in the previous section, the transition from hypoxia to a well-oxygenated state (i.e., reoxygenation) may be a period of vulnerability for breathing, as respiratory rhythmogenesis has a delayed recovery upon reoxygenation. Many biological systems exhibit an oxidative burst during reoxygenation (Granger and Kviety, 2015; Spranger et al., 1998) that can cause cellular injury (Granger and Kviety, 2015) and, in the context of the preBötC, affect stable rhythmogenesis. The oxidative burst results from one or more types of reactive oxygen species (ROS) being produced (Granger and Kviety, 2015; Halliwell, 1991; Li and Jackson, 2002). During reoxygenation, one type of ROS, superoxide anion, may be produced from the mitochondria (Bao et al., 2009; Gruber et al., 2013) or other cellular

processes (Bedard and Krause, 2007; Berry and Hare, 2004). Superoxide anion is rapidly converted, via superoxide dismutase, into hydrogen peroxide (H_2O_2) (Granger and Kvietyts, 2015). H_2O_2 is another type of ROS and can be further metabolized by catalase or other peroxidases (Granger and Kvietyts, 2015). Both superoxide anion and H_2O_2 have been implicated in cell signaling, but when they accumulate, can also be sources for oxidative stress (Granger and Kvietyts, 2015). For example, in presence of a transition metal, like Fe^{2+} , H_2O_2 is rapidly converted into the short-lived ROS, hydroxyl radical (Granger and Kvietyts, 2015). Unlike superoxide anion and H_2O_2 , hydroxyl radical does not act as a signaling molecule but rather, serves as a source of oxidative stress (Halliwell, 1991).

Acute H_2O_2 suppresses synaptic transmission in both the adult and neonatal hippocampus (Avshalumov and Rice, 2002; Garcia et al., 2011) yet H_2O_2 differentially affects rhythm generation from the preBötC (Garcia et al., 2011). Application of H_2O_2 to the in vitro slice triggers a biphasic change in the frequency of rhythmogenesis that is characterized by an initial depression followed by an augmentation phase. This response was remarkably similar to the stereotypical response of the isolated preBötC to reoxygenation from hypoxia and could be traced to individual neuronal responses to H_2O_2 . The initial depression of rhythmogenesis by H_2O_2 appears to be the result of oxidative stress, as hydroxyl radical, produced by co-application of Fe^{2+} and H_2O_2 (to produce hydroxyl radical) leading to monotonic depression in rhythmic frequency. By contrast, in the presence of the iron chelator, deferox-amine, H_2O_2 only causes augmentation of rhythmic frequency from the preBötC. The chelation of free iron minimized the conversion of H_2O_2 to hydroxyl radical, presumably unmasking the action that H_2O_2 has independent of hydroxyl radical. Indeed, optogenetic stimulation of microglia and hypoxia causes increases in extracellular H_2O_2 in ventrolateral medullary slices (Pardo-Pena et al., 2018). Thus, rhythmogenesis from the preBötC may be influenced by ROS in a species dependent fashion and such effects may have particular relevance to reoxygenation.

1.7. Intermittent hypoxia

While it may only take a single bout of hypoxia to destabilize breathing, intermittent oscillations in arterial O_2 are common to several conditions during neonatal life. To determine how such oscillations influence neonatal respiratory control, exposure to intermittent hypoxia has been used to experimentally model unstable breathing (Garcia et al., 2017, 2016; Peng et al., 2004; Prabhakar et al., 2007; Zanella et al., 2014). Studies in the preBötC have used both acute intermittent hypoxia and chronic intermittent hypoxia to understand the consequences of intermittent hypoxia versus hypoxia on inspiratory rhythmogenesis relevant to conditions where neonatal apneas are evident (Garcia et al., 2016; Zanella et al., 2014).

1.8. Acute intermittent hypoxia

Neonatal abstinence syndrome (NAS) is a condition where the neonatal infant experiences drug withdrawal due to maternal narcotic use or abuse during pregnancy (Kocherlakota, 2014; "Neonatal drug withdrawal. American Academy of Pediatrics Committee on Drugs, 1998). As many drugs readily pass from the maternal blood stream through the placenta to the fetus (Ross et al., 2015), illicit drug abuse during pregnancy can cause the fetus to

become addicted to the substance(s) used by the mother (Behnke et al., 2013). As a result, the sudden discontinuation of fetal exposure to substances that were used or abused by the mother during pregnancy commonly lead to withdrawal symptoms in the newborn and is associated with increased adrenergic tone (Oji-Mmuo et al., 2019) and disrupted serotonergic tone (Lunden and Kirby, 2013). Such departures from the normal neuromodulatory state could increase the risk of unstable breathing and oscillations in blood gases. Indeed, in premature infants suffering from NAS, intermittent hypoxemic episodes can worsen (Abu Jawdeh et al., 2017).

At the level of the preBötC, acute intermittent hypoxia (IH) produces the stereotypical responses of rhythmogenesis during each cycle of hypoxia and reoxygenation (Blitz and Ramirez, 2002; Zanella et al., 2014) and leads to long-lasting augmentation in the frequency of rhythmogenesis during the post-hypoxic period (Blitz and Ramirez, 2002; Camacho-Hernandez et al., 2018). Acute IH also has little impact on the transmission of the premotor rhythm originating from the preBötC and generated by the hypoglossal motor pool demonstrating the robustness of the network to repeated changes in oxygenation (Zanella et al., 2014). However, when combined with enhanced adrenergic tone, acute IH can cause long-lasting instabilities in inspiratory rhythmogenesis.

Enhanced adrenergic tone co-incident with acute IH leads to the emergence of post-hypoxic subnetwork bursting in the preBötC (Zanella et al., 2014). This subnetwork bursting in the premotor network fails to produce the motor bursts in the hypoglossal motor nucleus and subsequently, enhances irregularities both in the preBötC rhythm and inspiratory motor rhythm. This appears to be the result of functional remodeling that involves changes in glycinergic inhibition and activation of α_2 adrenergic receptors (Zanella et al., 2014). Thus, increased adrenergic tone and acute IH may serve to perpetuate perinatal respiratory instability causing the preBötC to enter into an unstable state where synaptic inhibition disrupts transmission from the premotor network to respiratory motor pools.

1.9. Chronic intermittent hypoxia

The occurrence of intermittent hypoxia during perinatal life is often experienced over the course of days and even weeks. For the majority of infants (< 30 weeks of gestation) suffering from apneas of prematurity, chronic intermittent hypoxemia associated with the condition does not diminish until after the 40th week of gestation (Eichenwald and Committee on, F., and Newborn, A. A. o. P., 2016). Perinatal exposure to chronic intermittent hypoxia (IH) enhances the neonatal ventilatory response to acute hypoxia (Garcia et al., 2016; Peng et al., 2004). The effects of chronic IH on neonatal ventilation involve both enhanced peripheral O_2 chemosensitivity of the carotid bodies (Peng et al., 2004; Prabhakar et al., 2007) and changed inspiratory rhythmogenesis from the preBötC (Garcia et al., 2017, 2016). Chronic IH impacts inspiratory rhythmogenesis in a state dependent manner. Following exposure to chronic IH, excitability during a network burst is reduced among individual inspiratory neurons and is co-incident with a loss of fidelity of individual neurons with the network rhythm (Garcia et al., 2016). These cellular effects on preBötC neurons result in increased short-term variability of both amplitude and period of the network rhythm and consequently increases transmission failure of the preBötC rhythm

to the hypoglossal motor pool in local respiratory circuit. Biochemically, chronic IH increases oxidative stress in the preBötC and irregular rhythmogenesis and transmission failure can be mitigated by administration of the antioxidant, manganese(III) tetrakis(1-methyl-4-pyridyl) porphyrin (MnTMPyP), during neonatal chronic IH exposure.

Neonatal CIH causes ROS-dependent endothelin signaling to augment the hypoxic response of the carotid bodies (Pawar et al., 2009). Similarly, in the adrenal medulla, neonatal chronic IH leads to a ROS-mediated suppression of multiple nicotinic receptor subunits which thereby impairs nicotinic receptor mediated activation of the adrenal medulla (Souvannakitti et al., 2010) and can last into adulthood (Souvannakitti et al., 2009). Thus, the role of chronic IH-dependent ROS in remodeling cellular and physiological activity does not appear to be a unique feature at the level of the preBötC but rather, is evident throughout the neonatal nervous system.

In adults, chronic IH causes oxidative stress that also potentiates the activity from the carotid bodies (Del Rio et al., 2010; Peng et al., 2006) and may be an upstream contributor to the pro-inflammatory state caused by chronic IH that promotes the potentiation of the carotid body (Del Rio et al., 2012) and the hypoxic central chemoreflex pathway (Oyarce & Iturriaga, 2018). Increased hypoxia inducible factor 1a signaling (HIF1a) in the nervous system is upstream of oxidative stress produced by chronic IH (Peng et al., 2006). Although it is unknown whether chronic IH causes increased HIF1a signaling in the preBötC, recent transcriptional analysis of neonatal preBötC neurons has revealed that HIF1a is expressed approximately three times higher in glutamatergic Dbx1 preBötC neurons than in non-Dbx1 neurons (Hayes et al., 2017). These observations suggest that chronic IH may indeed increase HIF1a signaling in the preBötC but this remains to be demonstrated. Should chronic IH increase HIF1a signaling in the preBötC, the role of such signaling remains to be determined. On one hand, HIF1a signaling may serve to be protective promoting survival during hypoxia, while on the other, increased HIF1a signaling due to oxidative stress may serve to promote a pro-oxidant state (Semenza and Prabhakar, 2007) to destabilize function in inspiratory brain circuit. However, the protective role and the pro-oxidant role for IH-dependent HIF1a signaling may not be mutually exclusive given the potential for reactive oxygen species to modulate rhythmogenesis (Garcia et al., 2011).

2. Conclusion

Neonatal breathing possesses the capability to respond to hypoxia but is often still vulnerable to disturbances that cause repeated oscillations in blood oxygen. Such destabilization can arise from a host of conditions, yet hypoxic challenge and unstable breathing must be overcome by the neonate in order to survive. While several parts of the nervous system contribute the control over breathing, inspiratory rhythm generation from the preBötC appears to emerge well-before full gestational age. The cellular and network properties of the inspiratory network normally permit a robust response to hypoxic challenge yet the transitions from well-oxygenated states to hypoxia and back reveal vulnerabilities in rhythm generation relevant to understanding conditions such as SIDS. Moreover, the response to recurrent hypoxia and reoxygenation, whether acute or chronic, reveals a hysteresis in rhythmogenesis from preBötC which may serve to perpetuate unstable

breathing when respiratory control is underdeveloped. Even if hypoxic insult during neonatal life does not result in sudden death, exposure to hypoxia early in life may have consequences that are not yet fully understood. This underscores the importance for continued the study of respiratory control and the premotor circuits that drive breathing. Pinpointing critical vulnerabilities in the control of breathing, whether to under-development or to pathophysiology may facilitate novel therapies to better mitigate or even prevent neonatal hypoxic insult and its negative consequences.

Funding

This work was supported by a grant from The BSD Office of Diversity & Inclusion at The University of Chicago awarded to AJG, by a NIH R01 NS10742101 awarded to AJG and by NIH P01 HL144454-01A1.

References

- Abu Jawdeh EG, Westgate PM, Pant A, Stacy AL, Mamilla D, Gabrani A, Giannone P, et al., 2017 Prenatal Opioid Exposure and Intermittent Hypoxemia in Preterm Infants: A Retrospective Assessment. *Front. Pediatr* 5, 253 10.3389/fped.2017.00253. [PubMed: 29270395]
- Alvaro RE, Rigatto H, 2016 Control of breathing in newborns In: Buonocore G, Bracci R, Weindling M (Eds.), *Neonatology: A Practical Approach to Neonatal Diseases*. Springer International Publishing, Cham, pp. 1–16.
- Anderson TM, Garcia AJ 3rd, Baertsch NA, Pollak J, Bloom JC, Wei AD, Ramirez JM, et al., 2016 A novel excitatory network for the control of breathing. *Nature* 536 (7614), 76–80. 10.1038/nature18944. [PubMed: 27462817]
- Anderson TM, Ramirez JM, 2017 Respiratory rhythm generation: triple oscillator hypothesis. *F1000Res* 6, 139 10.12688/f1000research.10193.1. [PubMed: 28299192]
- Avshalumov MV, Rice ME, 2002 NMDA receptor activation mediates hydrogen peroxide-induced pathophysiology in rat hippocampal slices. *J. Neurophysiol* 87 (6), 2896–2903. 10.1152/jn.2002.87.6.2896. [PubMed: 12037193]
- Bancalari E, Claure N, 2018 Respiratory instability and hypoxemia episodes in preterm infants. *Am. J. Perinatol* 35 (6), 534–536. 10.1055/s-0038-1637760. [PubMed: 29694990]
- Bao L, Avshalumov MV, Patel JC, Lee CR, Miller EW, Chang CJ, Rice ME, 2009 Mitochondria are the source of hydrogen peroxide for dynamic brain-cell signaling. *J. Neurosci* 29 (28), 9002–9010. 10.1523/JNEUROSCI.1706-09.2009. [PubMed: 19605638]
- Barkin RM, Hartley MR, Brooks JG, 1981 Influence of high altitude on sudden infant death syndrome. *Pediatrics* 68 (6), 891–892. [PubMed: 7322728]
- Barlow SM, Estep M, 2006 Central pattern generation and the motor infrastructure for suck, respiration, and speech. *J. Commun. Disord* 39 (5), 366–380. 10.1016/j.jcomdis.2006.06.011. [PubMed: 16876186]
- Bedard K, Krause KH, 2007 The NOX family of ROS-generating NADPH oxidases: physiology and pathophysiology. *Physiol. Rev* 87 (1), 245–313. 10.1152/physrev.00044.2005. [PubMed: 17237347]
- Behnke M, Smith VC, Committee on Substance, A, Committee on, F, Newborn, 2013 Prenatal substance abuse: short- and long-term effects on the exposed fetus. *Pediatrics* 131 (3), e1009–e1024. 10.1542/peds.2012-3931. [PubMed: 23439891]
- Bellingham MC, 1998 Driving respiration: the respiratory central pattern generator. *Clin. Exp. Pharmacol. Physiol* 25 (10), 847–856. [PubMed: 9784928]
- Berry CE, Hare JM, 2004 Xanthine oxidoreductase and cardiovascular disease: molecular mechanisms and pathophysiological implications. *J. Physiol. (Paris)* 555 (Pt 3), 589–606. 10.1113/jphysiol.2003.055913.
- Blitz DM, Ramirez JM, 2002 Long-term modulation of respiratory network activity following anoxia in vitro. *J. Neurophysiol* 87 (6), 2964–2971. 10.1152/jn.2002.87.6.2964. [PubMed: 12037199]

- Braman SS, 1995 The regulation of normal lung function. *Allergy Proc.* 16 (5), 223–226. [PubMed: 8566733]
- Butera RJ Jr., Rinzel J, Smith JC, 1999 Models of respiratory rhythm generation in the pre-Botzinger complex. I. Bursting pacemaker neurons. *J. Neurophysiol* 82 (1), 382–397. 10.1152/jn.1999.82.1.382. [PubMed: 10400966]
- Byard RW, 2018 The autopsy and pathology of sudden infant death syndrome In: Duncan JR, Byard RW (Eds.), *SIDS Sudden Infant and Early Childhood Death: The Past, the Present and the Future.* Adelaide (AU).
- Camacho-Hernandez NP, Lorea-Hernandez JJ, Pena-Ortega F, 2018 Microglial modulators reduce respiratory rhythm long-term facilitation in vitro. *Respir. Physiol. Neurobiol* 10.1016/j.resp.2018.07.012.
- Carroll MS, Ramirez JM, 2013 Cycle-by-cycle assembly of respiratory network activity is dynamic and stochastic. *J. Neurophysiol* 109 (2), 296–305. 10.1152/jn.00830.2011. [PubMed: 22993257]
- Carroll MS, Viemari JC, Ramirez JM, 2013 Patterns of inspiratory phase-dependent activity in the in vitro respiratory network. *J. Neurophysiol* 109 (2), 285–295. 10.1152/jn.00619.2012. [PubMed: 23076109]
- Cerpa VJ, Wu Y, Bravo E, Teran FA, Flynn RS, Richerson GB, 2017 Medullary 5-HT neurons: switch from tonic respiratory drive to chemoreception during postnatal development. *Neuroscience* 344, 1–14. 10.1016/j.neuroscience.2016.09.002. [PubMed: 27619736]
- Champagnat J, Morin-Surun MP, Bouvier J, Thoby-Brisson M, Fortin G, 2011 Prenatal development of central rhythm generation. *Respir. Physiol. Neurobiol* 178 (1), 146–155. 10.1016/j.resp.2011.04.013. [PubMed: 21527363]
- Chen J, Magnusson J, Karsenty G, Cummings KJ, 2013 Time- and age-dependent effects of serotonin on gasping and autoresuscitation in neonatal mice. *J Appl Physiol* (1985) 114 (12), 1668–1676. 10.1152/jappphysiol.00003.2013. [PubMed: 23558391]
- Chevalier M, De Sa R, Carroit L, Thoby-Brisson M, 2016a Mechanisms underlying adaptation of respiratory network activity to modulatory stimuli in the mouse embryo. *Neural Plast*, 2016, 3905257 10.1155/2016/3905257. [PubMed: 27239348]
- Chevalier M, Toporikova N, Simmers J, Thoby-Brisson M, 2016b Development of pacemaker properties and rhythmogenic mechanisms in the mouse embryonic respiratory network. *Elife* 5 10.7554/eLife.16125.
- Coles SK, Ernsberger P, Dick TE, 1998 Post-hypoxic frequency decline does not depend on alpha2-adrenergic receptors in the adult rat. *Brain Res.* 794 (2), 267–273. [PubMed: 9622648]
- Del Negro CA, Funk GD, Feldman JL, 2018 Breathing matters. *Nat. Rev. Neurosci* 19 (6), 351–367. 10.1038/s41583-018-0003-6. [PubMed: 29740175]
- Del Rio R, Moya EA, Iturriaga R, 2010 Carotid body and cardiorespiratory alterations in intermittent hypoxia: the oxidative link. *Eur. Respir. J* 36 (1), 143–150. 10.1183/09031936.00158109. [PubMed: 19996187]
- Del Rio R, Moya EA, Parga MJ, Madrid C, Iturriaga R, 2012 Carotid body inflammation and cardiorespiratory alterations in intermittent hypoxia. *Eur. Respir. J* 39 (6), 1492–1500. 10.1183/09031936.00141511. [PubMed: 22183481]
- Di Pasquale E, Monteau R, Hilaire G, 1994 Involvement of the rostral ventro-lateral medulla in respiratory rhythm genesis during the peri-natal period: an in vitro study in newborn and fetal rats. *Brain Res. Dev. Brain Res* 78 (2), 243–252. [PubMed: 8026078]
- Eichenwald EC, Committee on F, & Newborn A. A. o. P, 2016 Apnea of prematurity. *Pediatrics* 137 (1). 10.1542/peds.2015-3757.
- Lambert Erck, Parks AB, Cottengim SE,C, Faulkner M, Hauck FR, ShapiroMendoza CK, 2019 Sleep-related infant suffocation deaths attributable to Soft bedding, overlay, and wedging. *Pediatrics* 143 (5). 10.1542/peds.2018-3408.
- Ferguson AH, 2015 Ignored disease or diagnostic dustbin? Sudden infant death syndrome in the british context. *Soc. Hist. Med* 28 (3), 487–508. 10.1093/shm/hkv003. [PubMed: 26217070]
- Filiano JJ, Kinney HC, 1994 A perspective on neuropathologic findings in victims of the sudden infant death syndrome: the triple-risk model. *Biol. Neonate* 65 (3–4), 194–197. 10.1159/000244052. [PubMed: 8038282]

- Fujii M, Arata A, 2010 Adrenaline modulates on the respiratory network development. *Adv. Exp. Med. Biol* 669, 25–28. 10.1007/978-1-4419-5692-7_5. [PubMed: 20217314]
- Garcia AJ 3rd, Dashevskiy T, Khuu MA, Ramirez JM, 2017 Chronic intermittent hypoxia differentially impacts different states of inspiratory activity at the level of the preBotzinger complex. *Front. Physiol* 8, 571 10.3389/fphys.2017.00571. [PubMed: 28936176]
- Garcia AJ 3rd, Khan SA, Kumar GK, Prabhakar NR, Ramirez JM, 2011 Hydrogen peroxide differentially affects activity in the pre-Botzinger complex and hippocampus. *J. Neurophysiol* 106 (6), 3045–3055. 10.1152/jn.00550.2010. [PubMed: 21849609]
- Garcia AJ 3rd, Koschnitzky JE, Ramirez JM, 2013a The physiological determinants of sudden infant death syndrome. *Respir. Physiol. Neurobiol* 189 (2), 288–300. 10.1016/j.resp.2013.05.032. [PubMed: 23735486]
- Garcia AJ 3rd, Rotem-Kohavi N, Doi A, Ramirez JM, 2013b Post-hypoxic recovery of respiratory rhythm generation is gender dependent. *PLoS One* 8 (4), e60695 10.1371/journal.pone.0060695. [PubMed: 23593283]
- Garcia AJ 3rd, Zanella S, Dashevskiy T, Khan SA, Khuu MA, Prabhakar NR, Ramirez JM, 2016 Chronic intermittent hypoxia alters local respiratory circuit function at the level of the preBotzinger complex. *Front. Neurosci* 10, 4 10.3389/fnins.2016.00004. [PubMed: 26869872]
- Gauda EB, Cristofalo E, Nunez J, 2007 Peripheral arterial chemoreceptors and sudden infant death syndrome. *Respir. Physiol. Neurobiol* 157 (1), 162–170. 10.1016/j.resp.2007.02.016. [PubMed: 17446144]
- Gauda EB, Martin RJ, 2012 *Control of Breathing*. Elsevier, pp. 584–597.
- Givan SA, Cummings KJ, 2016 Intermittent severe hypoxia induces plasticity within serotonergic and catecholaminergic neurons in the neonatal rat ventrolateral medulla. *J Appl Physiol* (1985) 120 (11), 1277–1287. 10.1152/jappphysiol.00048.2016. [PubMed: 26968026]
- Granger DN, Kviety PR, 2015 Reperfusion injury and reactive oxygen species: the evolution of a concept. *Redox Biol.* 6, 524–551. 10.1016/j.redox.2015.08.020. [PubMed: 26484802]
- Grillner S, 2003 The motor infrastructure: from ion channels to neuronal networks. *Nat. Rev. Neurosci* 4 (7), 573–586. 10.1038/nrn1137. [PubMed: 12838332]
- Gross TL, Sokol RJ, Kwong MS, Wilson M, Kuhnert PM, 1983 Transient tachypnea of the newborn: the relationship to preterm delivery and significant neonatal morbidity. *Am. J. Obstet. Gynecol* 146 (3), 236–241. 10.1016/00029378(83)90742-1. [PubMed: 6859131]
- Gruber J, Fong S, Chen CB, Yoong S, Pastorin G, Schaffer S, Halliwell B, et al., 2013 Mitochondria-targeted antioxidants and metabolic modulators as pharmacological interventions to slow ageing. *Biotechnol. Adv* 31 (5), 563–592. 10.1016/j.biotechadv.2012.09.005. [PubMed: 23022622]
- Guyenet PG, Mulkey DK, 2010 Retrotrapezoid nucleus and parafacial respiratory group. *Respir. Physiol. Neurobiol* 173 (3), 244–255. 10.1016/j.resp.2010.02.005. [PubMed: 20188865]
- Guyenet PG, Mulkey DK, Stornetta RL, Bayliss DA, 2005 Regulation of ventral surface chemoreceptors by the central respiratory pattern generator. *J. Neurosci* 25 (39), 8938–8947. 10.1523/JNEUROSCI.2415-05.2005. [PubMed: 16192384]
- Halliwell B, 1991 Reactive oxygen species in living systems: source, biochemistry, and role in human disease. *Am. J. Med* 91 (3C), 14S–22S.
- Hayes JA, Kottick A, Picardo MCD, Halleran AD, Smith RD, Smith GD, Del Negro CA, et al., 2017 Transcriptome of neonatal preBotzinger complex neurones in Dbx1 reporter mice. *Sci. Rep* 7 (1), 8669 10.1038/s41598-01709418-4. [PubMed: 28819234]
- Hill AA, Garcia AJ 3rd, Zanella S, Upadhyaya R, Ramirez JM, 2011 Graded reductions in oxygenation evoke graded reconfiguration of the isolated respiratory network. *J. Neurophysiol* 105 (2), 625–639. 10.1152/jn.00237.2010. [PubMed: 21084689]
- Hillman NH, Kallapur SG, Jobe AH, 2012 Physiology of transition from intrauterine to extrauterine life. *Clin. Perinatol* 39 (4), 769–783. 10.1016/j.clp.2012.09.009. [PubMed: 23164177]
- Hodges MR, Wehner M, Aungst J, Smith JC, Richerson GB, 2009 Transgenic mice lacking serotonin neurons have severe apnea and high mortality during development. *J. Neurosci* 29 (33), 10341–10349. 10.1523/JNEUROSCI.1963-09.2009. [PubMed: 19692608]
- Hubbard JL, Shingleton HM, 1985 Sexual function of patients after cancer of the cervix treatment. *Clin. Obstet. Gynaecol* 12 (1), 247–264. [PubMed: 3888485]

- Janczewski WA, Feldman JL, 2006 Distinct rhythm generators for inspiration and expiration in the juvenile rat. *J. Physiol. (Paris)* 570 (Pt 2), 407–420. 10.1113/jphysiol.2005.098848.
- Kinney HC, Filiano JJ, White WF, 2001 Medullary serotonergic network deficiency in the sudden infant death syndrome: review of a 15-year study of a single dataset. *J. Neuropathol. Exp. Neurol* 60 (3), 228–247. [PubMed: 11245208]
- Kocherlakota P, 2014 Neonatal abstinence syndrome. *Pediatrics* 134 (2), e547–561. 10.1542/peds.2013-3524. [PubMed: 25070299]
- Koshiya N, Smith JC, 1999 Neuronal pacemaker for breathing visualized in vitro. *Nature* 400 (6742), 360–363. 10.1038/22540. [PubMed: 10432113]
- Li C, Jackson RM, 2002 Reactive species mechanisms of cellular hypoxia-reoxygenation injury. *Am. J. Physiol., Cell Physiol* 282 (2), C227–C241. 10.1152/ajpcell.00112.2001. [PubMed: 11788333]
- Lieske SP, Ramirez JM, 2006 Pattern-specific synaptic mechanisms in a multifunctional network. I. Effects of alterations in synapse strength. *J. Neurophysiol* 95 (3), 1323–1333. 10.1152/jn.00505.2004. [PubMed: 16492944]
- Lieske SP, Thoby-Brisson M, Telgkamp P, Ramirez JM, 2000 Reconfiguration of the neural network controlling multiple breathing patterns: eupnea, sighs and gasps [see comment]. *Nat. Neurosci* 3 (6), 600–607. 10.1038/75776. [PubMed: 10816317]
- Liu Q, Wong-Riley MT, 2005 Postnatal developmental expressions of neurotransmitters and receptors in various brain stem nuclei of rats. *J Appl Physiol* (1985) 98 (4), 1442–1457. 10.1152/jappphysiol.01301.2004. [PubMed: 15618314]
- Liu Q, Wong-Riley MT, 2006 Developmental changes in the expression of GABAA receptor subunits alpha1, alpha2, and alpha3 in brain stem nuclei of rats. *Brain Res.* 1098 (1), 129–138. 10.1016/j.brainres.2006.05.001. [PubMed: 16750519]
- Liu Q, Wong-Riley MT, 2012 Postnatal development of Na(+)-K(+)-2Cl(−) co-transporter 1 and K(+)-Cl(−) co-transporter 2 immunoreactivity in multiple brain stem respiratory nuclei of the rat. *Neuroscience* 210, 1–20. 10.1016/j.neuroscience.2012.03.018. [PubMed: 22441038]
- Lunden JW, Kirby LG, 2013 Opiate exposure and withdrawal dynamically regulate mRNA expression in the serotonergic dorsal raphe nucleus. *Neuroscience* 254, 160–172. 10.1016/j.neuroscience.2013.08.071. [PubMed: 24055683]
- Mage DT, Donner EM, 2004 The fifty percent male excess of infant respiratory mortality. *Acta Paediatr.* 93 (9), 1210–1215. [PubMed: 15384886]
- Mage DT, Donner EM, 2014 Is excess male infant mortality from sudden infant death syndrome and other respiratory diseases X-linked? *Acta Paediatr.* 103 (2), 188–193. 10.1111/apa.12482. [PubMed: 24164639]
- Mage DT, Donner M, 2009 A unifying theory for SIDS. *Int J Pediatr*, 2009, 368270 10.1155/2009/368270. [PubMed: 20049339]
- Martin RJ, DiFiore JM, Jana L, Davis RL, Miller MJ, Coles SK, Dick TE, 1998 Persistence of the biphasic ventilatory response to hypoxia in preterm infants. *J. Pediatr* 132 (6), 960–964. [PubMed: 9627586]
- Morgado-Valle C, Feldman JL, 2007 NMDA receptors in preBotzinger complex neurons can drive respiratory rhythm independent of AMPA receptors. *J. Physiol. (Paris)* 582 (Pt 1), 359–368. 10.1113/jphysiol.2007.130617.
- Mulkey DK, Wenker IC, 2011 Astrocyte chemoreceptors: mechanisms of H⁺ sensing by astrocytes in the retrotrapezoid nucleus and their possible contribution to respiratory drive. *Exp. Physiol* 96 (4), 400–406. 10.1113/expphysiol.2010.053140. [PubMed: 21169332]
- Neary MT, Mohun TJ, Breckenridge RA, 2013 A mouse model to study the link between hypoxia, long QT interval and sudden infant death syndrome. *Dis. Model. Mech* 6 (2), 503–507. 10.1242/dmm.010587. [PubMed: 22977222]
- Neonatal drug withdrawal. American Academy of Pediatrics Committee on Drugs, 1998 *Pediatrics* 101 (6), 1079–1088.
- Nieto-Posadas A, Flores-Martinez E, Lorea-Hernandez JJ, Rivera-Angulo AJ, PerezOrtega JE, Bargas J, Pena-Ortega F, 2014 Change in network connectivity during fictive-gasping generation in hypoxia: prevention by a metabolic intermediate. *Front. Physiol* 5, 265 10.3389/fphys.2014.00265. [PubMed: 25101002]

- Oji-Mmuo CN, Speer RR, Gardner FC, Marvin MM, Hozella AC, Doheny KK, 2019 Prenatal opioid exposure heightens sympathetic arousal and facial expressions of pain/distress in term neonates at 24–48 hours post birth. *J. Matern. Fetal. Neonatal. Med* 1–181. 10.1080/14767058.2019.1588876.
- Oku Y, Masumiya H, Okada Y, 2007 Postnatal developmental changes in activation profiles of the respiratory neuronal network in the rat ventral medulla. *J. Physiol. (Paris)* 585 (Pt 1), 175–186. 10.1113/jphysiol.2007.138180.
- Oyarce MP, Iturriaga R, 2018 Proinflammatory cytokines in the nucleus of the solitary tract of hypertensive rats exposed to chronic intermittent hypoxia. *Adv. Exp. Med. Biol* 1071, 69–74. 10.1007/978-3-319-91137-3_8. [PubMed: 30357735]
- Pagliardini S, Ren J, Greer JJ, 2003 Ontogeny of the pre-Botzinger complex in perinatal rats. *J. Neurosci* 23 (29), 9575–9584. [PubMed: 14573537]
- Pardo-Pena K, Lorea-Hernandez JJ, Camacho-Hernandez NP, Ordaz B, VillasanaSalazar B, Morales-Villagran A, Pena-Ortega F, 2018 Hydrogen peroxide extracellular concentration in the ventrolateral medulla and its increase in response to hypoxia in vitro: possible role of microglia. *Brain Res.* 1692, 87–99. 10.1016/j.brainres.2018.04.032. [PubMed: 29715442]
- Paton JF, Abdala AP, Koizumi H, Smith JC, St-John WM, 2006 Respiratory rhythm generation during gasping depends on persistent sodium current. *Nat. Neurosci* 9 (3), 311–313. 10.1038/nn1650. [PubMed: 16474390]
- Pawar A, Nanduri J, Yuan G, Khan SA, Wang N, Kumar GK, Prabhakar NR, 2009 Reactive oxygen species-dependent endothelin signaling is required for augmented hypoxic sensory response of the neonatal carotid body by intermittent hypoxia. *Am. J. Physiol. Regul. Integr. Comp. Physiol* 296 (3), R735–R742. 10.1152/ajpregu.90490.2008. [PubMed: 19109370]
- Pawar A, Peng YJ, Jacono FJ, Prabhakar NR, 2008 Comparative analysis of neonatal and adult rat carotid body responses to chronic intermittent hypoxia. *J Appl Physiol* (1985) 104 (5), 1287–1294. 10.1152/jappphysiol.00644.2007. [PubMed: 18187605]
- Pena F, Meza-Andrade R, Paez-Zayas V, Gonzalez-Marin MC, 2008 Gasping generation in developing Swiss-Webster mice in vitro and in vivo. *Neurochem. Res* 33 (8), 1492–1500. 10.1007/s11064-008-9616-x. [PubMed: 18273701]
- Pena F, Parkis MA, Tryba AK, Ramirez JM, 2004 Differential contribution of pacemaker properties to the generation of respiratory rhythms during normoxia and hypoxia. *Neuron* 43 (1), 105–117. 10.1016/j.neuron.2004.06.023. [PubMed: 15233921]
- Pena F, Ramirez JM, 2002 Endogenous activation of serotonin-2A receptors is required for respiratory rhythm generation in vitro. *J. Neurosci* 22 (24), 11055–11064. [PubMed: 12486201]
- Pena F, Ramirez JM, 2004 Substance P-mediated modulation of pacemaker properties in the mammalian respiratory network. *J. Neurosci* 24 (34), 7549–7556. 10.1523/JNEUROSCI.1871-04.2004. [PubMed: 15329402]
- Peng YJ, Rennison J, Prabhakar NR, 2004 Intermittent hypoxia augments carotid body and ventilatory response to hypoxia in neonatal rat pups. *J Appl Physiol* (1985) 97 (5), 2020–2025. 10.1152/jappphysiol.00876.2003. [PubMed: 15258129]
- Peng YJ, Yuan G, Ramakrishnan D, Sharma SD, Bosch-Marce M, Kumar GK, Prabhakar NR, et al., 2006 Heterozygous HIF-1 α deficiency impairs carotid body-mediated systemic responses and reactive oxygen species generation in mice exposed to intermittent hypoxia. *J. Physiol. (Paris)* 577 (Pt 2), 705–716. 10.1113/jphysiol.2006.114033.
- Pepper DR, Landauer RC, Kumar P, 1995 Postnatal development of CO₂-O₂ interaction in the rat carotid body in vitro. *J. Physiol. (Paris)* 485 (Pt 2), 531–541. 10.1113/jphysiol.1995.sp020749.
- Porzionato A, Macchi V, De Caro R, 2018 Central and peripheral chemoreceptors in sudden infant death syndrome. *J. Physiol. (Paris)* 596 (15), 3007–3019. 10.1113/JP274355.
- Prabhakar NR, Peng YJ, Kumar GK, Pawar A, 2007 Altered carotid body function by intermittent hypoxia in neonates and adults: relevance to recurrent apneas. *Respir. Physiol. Neurobiol* 157 (1), 148–153. 10.1016/j.resp.2006.12.009. [PubMed: 17317339]
- Ramirez JM, Doi A, Garcia AJ 3rd, Elsen FP, Koch H, Wei AD, 2012 The cellular building blocks of breathing. *Compr. Physiol* 2 (4), 2683–2731. 10.1002/cphy.c110033. [PubMed: 23720262]

- Ramirez JM, Koch H, Garcia AJ 3rd, Doi A, Zanella S, 2011 The role of spiking and bursting pacemakers in the neuronal control of breathing. *J. Biol. Phys* 37 (3), 241–261. 10.1007/s10867-011-9214-z. [PubMed: 22654176]
- Ramirez JM, Ramirez SC, Anderson TM, 2018 Sudden infant death syndrome, sleep, and the physiology and pathophysiology of the respiratory network In: Duncan JR, Byard RW (Eds.), *SIDS Sudden Infant and Early Childhood Death: The Past, the Present and the Future*. Adelaide (AU)..
- Ramirez JM, Tryba AK, Pena F, 2004 Pacemaker neurons and neuronal networks: an integrative view. *Curr. Opin. Neurobiol* 14 (6), 665–674. 10.1016/j.conb.2004.10.011. [PubMed: 15582367]
- Reuter S, Moser C, Baack M, 2014 Respiratory distress in the newborn. *Pediatr. Rev* 35 (10), 417–428. 10.1542/pir.35-10-417. quiz 429. [PubMed: 25274969]
- Rigatto H, Brady JP, 1972 Periodic breathing and apnea in preterm infants. II. Hypoxia as a primary event. *Pediatrics* 50 (2), 219–228. [PubMed: 5045351]
- Rigatto H, Brady JP, de la Torre Verduzco R, 1975 Chemoreceptor reflexes in preterm infants: I. The effect of gestational and postnatal age on the ventilatory response to inhalation of 100% and 15% oxygen. *Pediatrics* 55 (5), 604–613. [PubMed: 1128986]
- Ross EJ, Graham DL, Money KM, Stanwood GD, 2015 Developmental consequences of fetal exposure to drugs: what we know and what we still must learn. *Neuropsychopharmacology* 40 (1), 61–87. 10.1038/npp.2014.147. [PubMed: 24938210]
- Semenza GL, Prabhakar NR, 2007 HIF-1-dependent respiratory, cardiovascular, and redox responses to chronic intermittent hypoxia. *Antioxid. Redox Signal* 9 (9), 1391–1396. 10.1089/ars.2007.1691. [PubMed: 17627473]
- Shao XM, Ge Q, Feldman JL, 2003 Modulation of AMPA receptors by cAMP-dependent protein kinase in preBotzinger complex inspiratory neurons regulates respiratory rhythm in the rat. *J. Physiol. (Paris)* 547 (Pt 2), 543–553. 10.1113/jphysiol.2002.031005.
- Smith JC, Ellenberger HH, Ballanyi K, Richter DW, Feldman JL, 1991 Pre-Botzinger complex: a brainstem region that may generate respiratory rhythm in mammals. *Science* 254 (5032), 726–729. [PubMed: 1683005]
- Souvannakitti D, Kumar GK, Fox A, Prabhakar NR, 2009 Neonatal intermittent hypoxia leads to long-lasting facilitation of acute hypoxia-evoked catecholamine secretion from rat chromaffin cells. *J. Neurophysiol* 101 (6), 2837–2846. 10.1152/jn.00036.2009. [PubMed: 19339466]
- Souvannakitti D, Kuri B, Yuan G, Pawar A, Kumar GK, Smith C, Prabhakar NR, et al., 2010 Neonatal intermittent hypoxia impairs neuronal nicotinic receptor expression and function in adrenal chromaffin cells. *Am. J. Physiol., Cell Physiol* 299 (2), C381–C388. 10.1152/ajpcell.00159.2010.10.1152/ajpcell.00530.2009. [PubMed: 20664070]
- Spranger M, Kiprianova I, Krempien S, Schwab S, 1998 Reoxygenation increases the release of reactive oxygen intermediates in murine microglia. *J. Cereb. Blood Flow Metab* 18 (6), 670–674. 10.1097/00004647-199806000-00009. [PubMed: 9626191]
- Stornetta RL, Moreira TS, Takakura AC, Kang BJ, Chang DA, West GH, Guyenet PG, et al., 2006 Expression of Phox2b by brainstem neurons involved in chemosensory integration in the adult rat. *J. Neurosci* 26 (40), 10305–10314. 10.1523/JNEUROSCI.2917-06.2006. [PubMed: 17021186]
- Stunden CE, Filosa JA, Garcia AJ, Dean JB, Putnam RW, 2001 Development of in vivo ventilatory and single chemosensitive neuron responses to hypercapnia in rats. *Respir. Physiol* 127 (2–3), 135–155. [PubMed: 11504586]
- Thach B, 2008 Tragic and sudden death. Potential and proven mechanisms causing sudden infant death syndrome. *EMBO Rep.* 9 (2), 114–118. 10.1038/sj.embor.7401163. [PubMed: 18246101]
- Thoby-Brisson M, Karlen M, Wu N, Charnay P, Champagnat J, Fortin G, 2009 Genetic identification of an embryonic parafacial oscillator coupling to the preBotzinger complex. *Nat. Neurosci* 12 (8), 1028–1035. 10.1038/nn.2354. [PubMed: 19578380]
- Thoby-Brisson M, Trinh JB, Champagnat J, Fortin G, 2005 Emergence of the pre-Botzinger respiratory rhythm generator in the mouse embryo. *J. Neurosci* 25 (17), 4307–4318. 10.1523/JNEUROSCI.0551-05.2005. [PubMed: 15858057]

- Tree K, Viemari JC, Cayetanot F, Peyronnet J, 2016 Growth restriction induced by chronic prenatal hypoxia affects breathing rhythm and its pontine catecholaminergic modulation. *J. Neurophysiol* 116 (4), 1654–1662. 10.1152/jn.00869.2015. [PubMed: 27486108]
- Tryba AK, Pena F, Ramirez JM, 2006 Gasping activity in vitro: a rhythm dependent on 5-HT_{2A} receptors. *J. Neurosci* 26 (10), 2623–2634. 10.1523/JNEUROSCI.4186-05.2006. [PubMed: 16525041]
- Tupal S, Huang WH, Picardo MC, Ling GY, Del Negro CA, Zoghbi HY, Gray PA, 2014 Atoh1-dependent rhombic lip neurons are required for temporal delay between independent respiratory oscillators in embryonic mice. *Elife* 3, e02265 10.7554/eLife.02265. [PubMed: 24842997]
- Viemari JC, Burnet H, Bevengut M, Hilaire G, 2003 Perinatal maturation of the mouse respiratory rhythm-generator: in vivo and in vitro studies. *Eur. J. Neurosci* 17 (6), 1233–1244. [PubMed: 12670311]
- Viemari JC, Garcia AJ 3rd, Doi A, Ramirez JM, 2011 Activation of alpha-2 noradrenergic receptors is critical for the generation of fictive eupnea and fictive gasping inspiratory activities in mammals in vitro. *Eur. J. Neurosci* 33 (12), 2228–2237. 10.1111/j.1460-9568.2011.07706.x. [PubMed: 21615559]
- Viemari JC, Ramirez JM, 2006 Norepinephrine differentially modulates different types of respiratory pacemaker and nonpacemaker neurons. *J. Neurophysiol* 95 (4), 2070–2082. 10.1152/jn.01308.2005. [PubMed: 16394066]
- Waggener TB, Southall DP, Scott LA, 1990 Analysis of breathing patterns in a prospective population of term infants does not predict susceptibility to sudden infant death syndrome. *Pediatr. Res* 27 (2), 113–117. 10.1203/00006450199002000-00002. [PubMed: 2314938]
- Wenker IC, Kreneisz O, Nishiyama A, Mulkey DK, 2010 Astrocytes in the retrotrapezoid nucleus sense H⁺ by inhibition of a Kir4.1-Kir5.1-like current and may contribute to chemoreception by a purinergic mechanism. *J. Neurophysiol* 104 (6), 3042–3052. 10.1152/jn.00544.2010. [PubMed: 20926613]
- Wong-Riley MT, Liu Q, 2005 Neurochemical development of brain stem nuclei involved in the control of respiration. *Respir. Physiol. Neurobiol* 149 (1–3), 83–98. 10.1016/j.resp.2005.01.011. [PubMed: 16203213]
- Wu M, Haxhiu MA, Johnson SM, 2005 Hypercapnic and hypoxic responses require intact neural transmission from the pre-Botzinger complex. *Respir. Physiol. Neurobiol* 146 (1), 33–46. 10.1016/j.resp.2004.11.005. [PubMed: 15733777]
- Yangzom Y, Qian L, Shan M, La Y, Meiduo D, Hu X, Zetterstrom R, et al., 2008 Outcome of hospital deliveries of women living at high altitude: a study from Lhasa in Tibet. *Acta Paediatr.* 97 (3), 317–321. 10.1111/j.1651-2227.2008.00628.x. [PubMed: 18298779]
- Young JO, Geurts A, Hodges MR, Cummings KJ, 2017 Active sleep unmasks apnea and delayed arousal in infant rat pups lacking central serotonin. *J Appl Physiol* (1985) 123 (4), 825–834. 10.1152/jappphysiol.00439.2017. [PubMed: 28775068]
- Zanella S, Doi A, Garcia AJ 3rd, Elsen F, Kirsch S, Wei AD, Ramirez JM, 2014 When norepinephrine becomes a driver of breathing irregularities: how intermittent hypoxia fundamentally alters the modulatory response of the respiratory network. *J. Neurosci* 34 (1), 36–50. 10.1523/JNEUROSCI.3644-12.2014. [PubMed: 24381266]

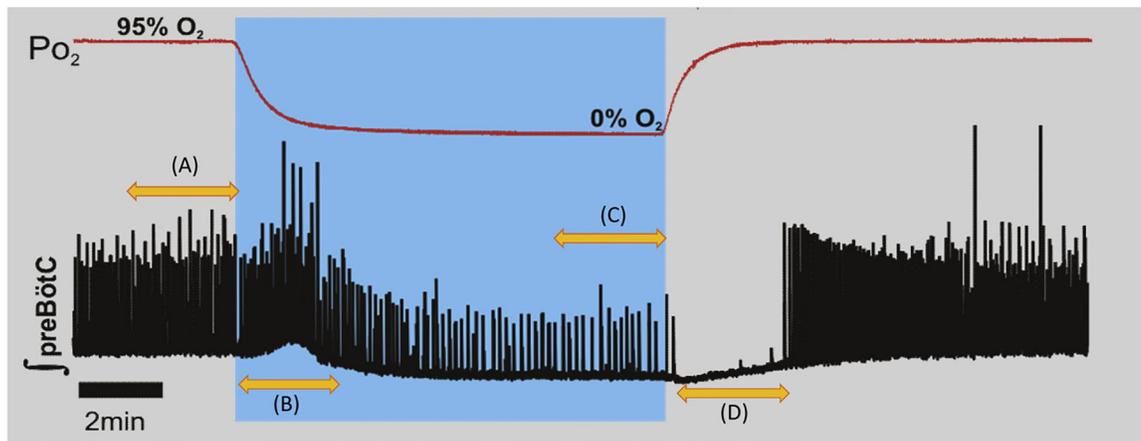


Fig. 1.

The stereotypical response of the isolated preBötC to hypoxia and reoxygenation. (A) The in vitro preBötC generates a spontaneous rhythm in well-oxygenated conditions (95% O₂ 5% CO₂). (B) Switching from to 95% N₂ 5% CO₂ produces hypoxic conditions as oxygen can still be measured in the circulating media (see Hill et al., 2011). During the initial period of hypoxic exposure, the in vitro rhythm increases in frequency and begins to generate augmented bursts. (C) Later during hypoxia augmented bursting ceases and rhythmogenesis slows. (D) Upon reoxygenation, the preBötC rhythm is initially suppressed delaying post-hypoxic recovery.